and aggregates. The preparation to be examined complies with the test if, in the chromatogram obtained with the test solution:

- **relative retention**: for the monomer and for the dimer, the relative retention to the corresponding peak in the chromatogram obtained with the reference solution is $1 \pm 0.02$;
- **peak area**: the sum of the peak areas of the monomer and the dimer represent not less than 85 per cent of the total area of the chromatogram and the sum of the peak areas of polymers and aggregates represents not more than 10 per cent of the total area of the chromatogram.

### Anti-A and anti-B haemagglutinins (2.6.20)

If human normal immunoglobulin is intended for subcutaneous administration, carry out the tests for anti-A and anti-B haemagglutinins. Dilute the preparation to be examined to an immunoglobulin concentration of 30 g/l before preparing the dilutions to be used in the test. The 64-fold dilutions do not show agglutination.

### Anti-D antibodies (2.6.26)

If human normal immunoglobulin is intended for subcutaneous administration, it complies with the test for anti-D antibodies in human immunoglobulin for intravenous administration.

### Antibody to hepatitis B surface antigen

Not less than 0.5 IU/g of immunoglobulin, determined by a suitable immunochemical method (2.7.1).

### Antibody to hepatitis A virus

If intended for use in the prophylaxis of hepatitis A, it complies with the following additional requirement. Determine the antibody content by comparison with a reference preparation calibrated in International Units, using an immunoassay of suitable sensitivity and specificity (2.7.1).

The International Unit is the activity contained in a stated amount of the International Standard for anti-hepatitis A immunoglobulin. The equivalence in International Units of the International Standard is stated by the World Health Organisation.

### Human hepatitis A immunoglobulin

BRP is calibrated in International Units by comparison with the International Standard.

The stated potency is not less than 100 IU/ml. The estimated potency is not less than the stated potency. The confidence limits ($P = 0.95$) of the estimated potency are not less than 80 per cent and not more than 125 per cent.

### Water

Determined by a suitable method, such as the semi-micro determination of water (2.5.12), loss on drying (2.2.32) or near infrared spectrophotometry (2.2.40), the water content is within the limits approved by the competent authority.

### Sterility (2.6.1)

It complies with the test for sterility.

### Pyrogens (2.6.8)

It complies with the test for pyrogens. Inject 1 ml per kilogram of the rabbit’s mass.

### STORAGE

For the liquid preparation, in a colourless glass container, protected from light. For the freeze-dried preparation, in an airtight colourless glass container, protected from light.

### LABELLING

The label states:

- for liquid preparations, the volume of the preparation in the container and the protein content expressed in grams per litre;
- for freeze-dried preparations, the quantity of protein in the container;
- the route of administration;
- for freeze-dried preparations, the name or composition and the volume of the reconstituting liquid to be added;
- where applicable, that the preparation is suitable for use in the prophylaxis of hepatitis A infection;
- where applicable, the anti-hepatitis A virus activity in International Units per millilitre;
- where applicable, the name and amount of antimicrobial preservative in the preparation.

**07/2008:0853**

### HUMAN PLASMA FOR FRACTIONATION

#### Plasma humanum ad separationem

**DEFINITION**

Human plasma for fractionation is the liquid part of human blood remaining after separation of the cellular elements from blood collected in a receptacle containing an anticoagulant, or separated by continuous filtration or centrifugation of anticoagulated blood in an apheresis procedure; it is intended for the manufacture of plasma-derived products.

#### PRODUCTION

**DONORS**

Only a carefully selected, healthy donor who, as far as can be ascertained after medical examination, laboratory blood tests and a study of the donor’s medical history, is free from detectable agents of infection transmissible by plasma-derived products may be used. Recommendations in this field are made by the Council of Europe [Recommendation No. R (95) 15 on the preparation, use and quality assurance of blood components, or subsequent revision]; a directive of the European Union also deals with the matter: Commission Directive 2004/33/EC of 22 March 2004 implementing Directive 2002/98/EC of the European Parliament and of the Council as regards certain technical requirements for blood and blood components.

### Immunisation of donors

Immunisation of donors to obtain immunoglobulins with specific activities may be carried out when sufficient supplies of material of suitable quality cannot be obtained from naturally immunised donors. Recommendations for such immunisations are formulated by the World Health Organisation (Requirements for the collection, processing and quality control of blood, blood components and plasma derivatives, WHO Technical Report Series, No. 840, 1994 or subsequent revision).

### Records

Records of donors and donations made are kept in such a way that, while maintaining the required degree of confidentiality concerning the donor’s identity, the origin of each donation in a plasma pool and the results of the corresponding acceptance procedures and laboratory tests can be traced.

### Laboratory tests

Laboratory tests are carried out for each donation to detect the following viral markers:

1. antibodies against human immunodeficiency virus 1 (anti-HIV-1);
2. antibodies against human immunodeficiency virus 2 (anti-HIV-2);
3. hepatitis B surface antigen (HBsAg);
4. antibodies against hepatitis C virus (anti-HCV).

The test methods used are of suitable sensitivity and specificity and comply with the regulations in force. If a
repeat-reactive result is found in any of these tests, the donation is not accepted.

**INDIVIDUAL PLASMA UNITS**

The plasma is prepared by a method that removes cells and cell debris as completely as possible. Whether prepared from whole blood or by plasmapheresis, the plasma is separated from the cells by a method designed to prevent the introduction of micro-organisms. No antibacterial or antifungal agent is added to the plasma. The containers comply with the requirements for glass containers (2.2.1) or for plastic containers for blood and blood components (3.2.3). The containers are closed so as to prevent any possibility of contamination.

If 2 or more units are pooled prior to freezing, the operations are carried out using sterile connecting devices or under aseptic conditions and using containers that have not previously been used.

When obtained by plasmapheresis or from whole blood (after separation from cellular elements), plasma intended for the recovery of proteins that are labile in plasma is frozen within 24 h of collection by cooling rapidly in conditions validated to ensure that a temperature of −25 °C or below is attained at the core of each plasma unit within 12 h of placing in the freezing apparatus.

When obtained by plasmapheresis, plasma intended solely for the recovery of proteins that are not labile in plasma is frozen by cooling rapidly in a chamber at −20 °C or below, as soon as possible and at the latest within 24 h of collection.

When obtained from whole blood, plasma intended solely for the recovery of proteins that are not labile in plasma is separated from cellular elements and frozen in a chamber at −20 °C or below as soon as possible and at the latest within 72 h of collection.

It is not intended that the determination of total protein and factor VIII shown below be carried out on each unit of plasma. They are rather given as guidelines for good manufacturing practice, the test for factor VIII being relevant for plasma intended for use in the preparation of concentrates of labile proteins.

The total protein content of a unit of plasma depends on the serum protein content of the donor and the degree of dilution inherent in the donation procedure. When plasma is obtained from a suitable donor and using the intended proportion of anticoagulant solution, a total protein content complying with the limit of 50 g/l is obtained. If a volume of blood or plasma smaller than intended is collected into the anticoagulant solution, the resulting plasma is not necessarily unsuitable for pooling for fractionation. The aim of good manufacturing practice must be to achieve the prescribed limit for all normal donations.

Preservation of factor VIII in the donation depends on the collection procedure and the subsequent handling of the blood and plasma. With good practice, 0.7 IU/ml can usually be achieved, but units of plasma with a lower activity may still be suitable for use in the production of coagulation factor concentrates. The aim of good manufacturing practice is to conserve labile proteins as much as possible.

**Total protein**. Carry out the test using a pool of not fewer than 10 units. Dilute the pool with a 9 g/l solution of sodium chloride R to obtain a solution containing about 15 mg of protein in 2 ml. To 2.0 ml of this solution in a round-bottomed centrifuge tube add 2 ml of a 75 g/l solution of sodium molybdate R and 2 ml of a mixture of 1 volume of nitrogen-free sulphuric acid R and 30 volumes of water R. Shake, centrifuge for 5 min, decant the supernatant liquid and allow the inverted tube to drain on filter paper.

Determine the nitrogen in the residue by the method of sulphuric acid digestion (2.5.9) and calculate the protein content by multiplying the quantity of nitrogen by 6.25. The total protein content is not less than 50 g/l.

**Factor VIII**. Carry out the test using a pool of not fewer than 10 units. Thaw the samples to be examined, if necessary, at 37 °C. Carry out the assay of factor VIII (2.7.4), using a reference plasma calibrated against the International Standard for human coagulation factor VIII in plasma. The activity is not less than 0.7 IU/ml.

**STORAGE AND TRANSPORT**

Frozen plasma is stored and transported in conditions designed to maintain the temperature at or below −20 °C; for accidental reasons, the storage temperature may rise above −20 °C on one or more occasions during storage and transport but the plasma is nevertheless considered suitable for fractionation if all the following conditions are fulfilled:

- the total period of time during which the temperature exceeds −20 °C does not exceed 72 h;
- the temperature does not exceed −15 °C on more than one occasion;
- the temperature at no time exceeds −5 °C.

**POOLED PLASMA**

During the manufacture of plasma products, the first homogeneous pool of plasma (for example, after removal of cryoprecipitate) is tested for HBsAg and for HIV antibodies using test methods of suitable sensitivity and specificity; the pool must give negative results in these tests.

The plasma pool is also tested for hepatitis C virus RNA using a validated nucleic acid amplification technique (2.6.21). A positive control with 100 IU/ml of hepatitis C virus RNA and, to test for inhibitors, an internal control prepared by addition of a suitable marker to a sample of the plasma pool are included in the test. The test is invalid if the positive control is non-reactive or if the result obtained with the internal control indicates the presence of inhibitors. The plasma pool complies with the test if it is found non-reactive for hepatitis C virus RNA.

**Hepatitis C virus RNA for NAT testing BRP** is suitable for use as a positive control.

**CHARACTERS**

Before freezing, a clear to slightly turbid liquid without visible signs of haemolysis; it may vary in colour from light yellow to green.

**LABELLING**

The label enables each individual unit to be traced to a specific donor.